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# $\beta_2$ Integrins As Regulators of Dendritic Cell, Monocyte, and Macrophage Function

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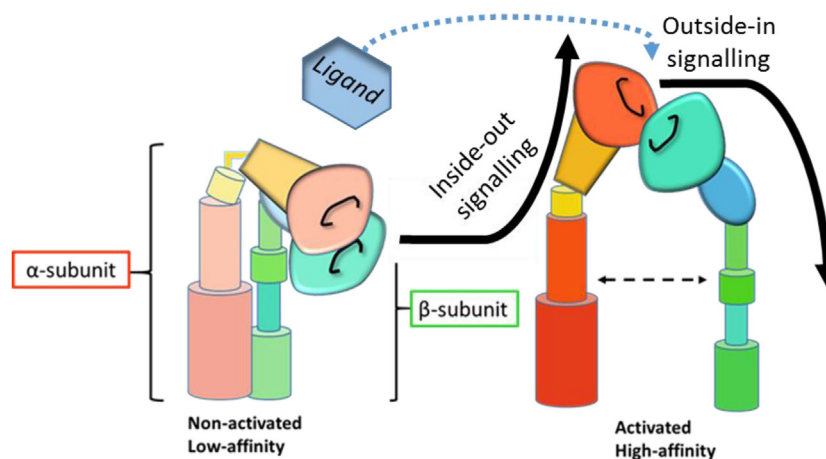
Emerging evidence suggests that the  $\beta_2$  integrin family of adhesion molecules have an important role in suppressing immune activation and inflammation.  $\beta_2$  integrins are important adhesion and signaling molecules that are exclusively expressed on leukocytes. The four  $\beta_2$  integrins (CD11a, CD11b, CD11c, and CD11d paired with the  $\beta_2$  chain CD18) play important roles in regulating three key aspects of immune cell function: recruitment to sites of inflammation; cell-cell contact formation; and downstream effects on cellular signaling. Through these three processes,  $\beta_2$  integrins both contribute to and regulate immune responses. This review explores the pro- and anti-inflammatory effects of  $\beta_2$  integrins in monocytes, macrophages, and dendritic cells and how they influence the outcome of immune responses. We furthermore discuss how imbalances in  $\beta_2$  integrin function can have far-reaching effects on mounting appropriate immune responses, potentially influencing the development and progression of autoimmune and inflammatory diseases. Therapeutic targeting of  $\beta_2$  integrins, therefore, holds enormous potential in exploring treatment options for a variety of inflammatory conditions.

**Keywords:**  $\beta_2$  integrins, CD11/CD18, dendritic cells monocytes and macrophages, immune regulation, autoimmunity

## INTRODUCTION

The integrin family of proteins is comprised of 24 heterodimeric transmembrane adhesion receptors. Each integrin is formed through the non-covalent association of 1  $\alpha$ -subunit and 1  $\beta$ -subunit; currently, 16  $\alpha$ -subunits and 8  $\beta$ -subunits have been identified. Their expression on virtually all human cells and their complex signaling mechanisms explain their wide variety of biological roles, including blood clotting, cell adhesion, and migration.

Due to their extensive importance in biological systems, elucidating integrin signaling and receptor function has been of great interest since their characterization as adhesion molecules over 30 years ago. Integrins are important signaling proteins that mediate interactions of the cell with extracellular matrix proteins and with other cells via cell-surface ligands. Integrins exist in a continuum between a folded inactive form with low affinity for their ligand and an extended high affinity conformation (1), although even bent integrins are able to bind ligand in rare instances (2). As immune cell adhesion and extravasation into lymph nodes and tissues forms part of initiating an effective immune response,  $\beta_2$  integrin conformation on the surface of leukocytes needs to be tightly regulated.  $\beta_2$  integrins on the surface of circulating leukocytes tend, therefore, to be largely inactive (2) until inside-out and outside-in signaling trigger integrin-mediated adhesion and extravasation into tissue (Figure 1).



**FIGURE 1** | Schematic representation of integrin activation and signaling. Inside-out signaling induces a conformational change in the integrin to the active, high affinity state. Upon ligand binding, active integrins then transmit outside-in signals and downstream signaling cascades. [Adapted from Byron et al. (3), with permission from the *Journal of Cell Science*].

Inside-out signaling modifies how cells interact *with* their environment by facilitating receptor affinity and avidity (4) to allow binding to extracellular ligands. Outside-in signaling, on the other hand, mediates intracellular events in response *to* their environment by eliciting downstream signaling cascades in response to receptor occupation. The complex details of integrin signaling are reviewed elsewhere (5, 6) and are beyond the scope of this review. Briefly, inside-out signaling is mediated by talin (7) and kindlin (8, 9) binding to the intracellular domain of the  $\beta_2$  subunit, a process initiated by chemokine receptor or Toll-like receptor (TLR) engagement (10, 11), which results in a conformational change in the integrin from a low-affinity to a high-affinity state. Outside-in signaling is then initiated by ligand binding to high-affinity integrin receptors (Figure 1). Downstream signaling events mediate the formation of focal complexes and adhesions through rearrangement of the actin cytoskeleton. The relative importance of affinity and avidity on integrin signaling and function is heavily debated (12, 13), but dynamic interaction between these processes and both inside-out and outside-in signaling seems likely (14).

$\beta_2$  integrins are the focus of this review, as they are exclusively found on leukocytes and therefore of particular importance for the immune system. They mediate cell recruitment into lymphoid organs and inflamed tissues by facilitating firm leukocyte arrest on endothelial cells and extravasation after cell rolling (15); cellular interactions between leukocytes including immunological synapse formation (16); and intracellular signaling cascades that influence cytoskeletal rearrangement, activation, proliferation and impact on cellular responses to TLRs. Importantly, through these three processes,  $\beta_2$  integrins can have either pro-inflammatory or anti-inflammatory outcomes. The  $\beta_2$  integrin subunit (CD18) can pair with one of four  $\alpha$ -subunits ( $\alpha_L$ —CD11a,  $\alpha_M$ —CD11b,  $\alpha_X$ —CD11c, and  $\alpha_D$ —CD11d), forming leukocyte function-associated antigen-1, Mac1/CR3 (macrophage-1 antigen, complement receptor 3), P150,95/CR4 (complement receptor 4), and CD18/CD11d, respectively (Figure 2). For consistency, this

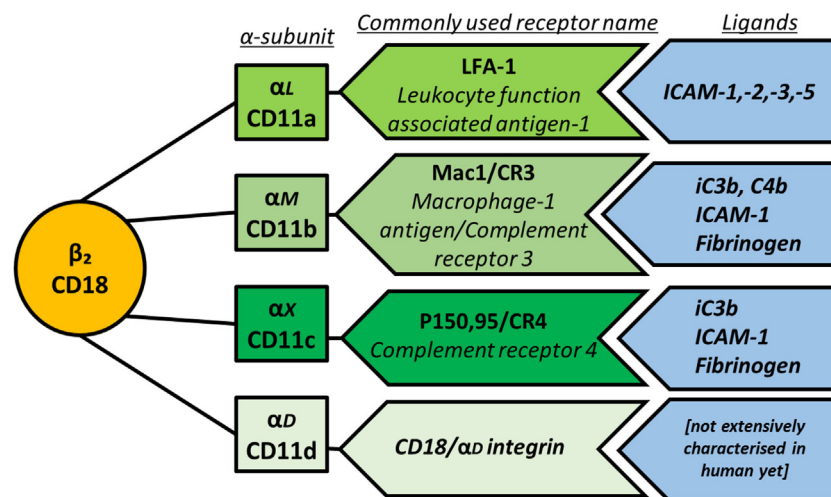
review will utilize only the CD nomenclature. Both function and cell-specific expression of  $\beta_2$  integrins vary according to the  $\alpha$ -subunit involved.

The main ligands for the  $\beta_2$  integrin family members are outlined in Figure 2. Briefly, CD11a binds to intracellular adhesion molecule-1 (ICAM-1), -2, -3, and -5, which are expressed by a variety of cells including leukocytes and endothelial cells, thereby mediating leukocyte recruitment to lymph nodes and sites of inflammation as well as cell-cell adhesion. CD11b binds the complement proteins iC3b and C4b with high affinity, mediating phagocytosis of complement-coated particles but can also bind ICAM-1, fibrinogen, and more than 40 other ligands (17). The sequence of CD11c is very close to that of CD11b, and indeed CD11c binds several of the same ligands including iC3b, ICAM-1, and fibrinogen. Multi-ligand binding capacity of CD11d is proposed to largely overlap with CD11b and includes ECM-associated proteins fibronectin, fibrinogen, vitronectin, Cyr61, and plasminogen (18).

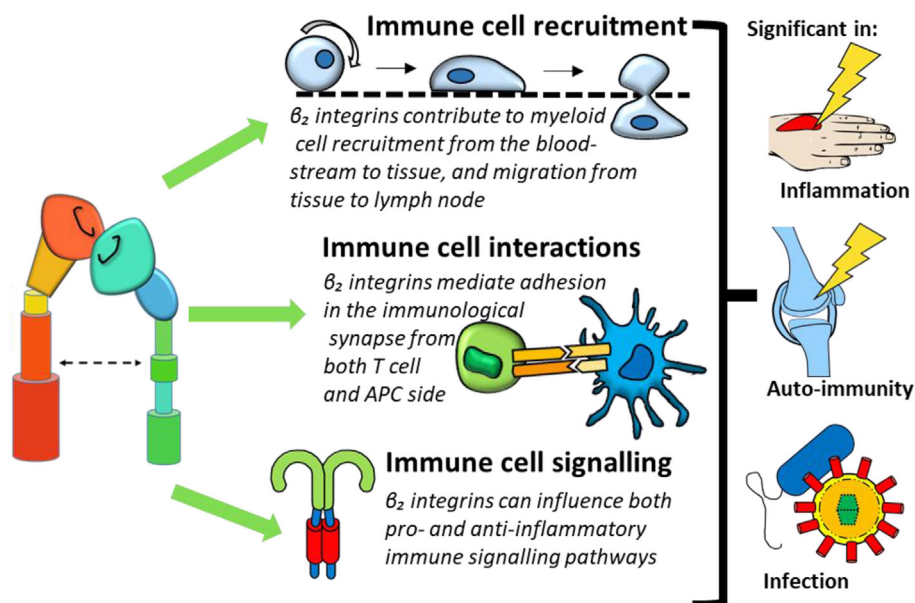
This review will provide an overview of  $\beta_2$  integrin expression on monocytes, macrophages and DCs, before exploring the paradoxical pro-inflammatory and regulatory roles of  $\beta_2$  integrins in immune regulation in three key aspects of immune function: recruitment and migration, cellular interactions, and downstream cell signaling (Figure 3). We will furthermore review how dysregulated integrin signaling could contribute to inflammatory and autoimmune conditions and introduce the therapeutic potential of targeting  $\beta_2$  integrins.

## EXPRESSION OF $\beta_2$ INTEGRIN SUBUNITS BY DENDRITIC CELLS (DCs), MONOCYTES, AND MACROPHAGES

The expression of  $\beta_2$  integrin subunits varies in different leukocyte subsets and between mice and humans. In general terms, CD11a is expressed on all leukocytes at varying levels, while CD11b,



**FIGURE 2** | Schematic representation of  $\beta_2$  integrin subunit pairing, depicting the  $\beta$ -subunit CD18 as the common subunit non-covalently associating with one of four  $\alpha$ -subunits. The main ligands for each integrin are also shown.



**FIGURE 3** |  $\beta_2$  integrin involvement in immune cell function can be categorized into three processes: immune cell recruitment, immune cell interactions, and immune cell signaling. Dysregulation of these functions could contribute to conditions such as inflammation, immunity, and infection.

CD11c, and CD11d are predominantly expressed by monocytes, macrophages and DCs. Specifically, in humans, monocytes express all four  $\beta_2$  integrin-associated alpha subunits (CD11a, CD11b, CD11c, and CD11d) with CD11a and CD11b expression greater than CD11c (19, 20); macrophages express CD11a and CD11b at lower levels than monocytes together with CD11c at similar levels to monocytes (21); while DCs mainly express CD11c together with CD11a, though some DC subsets also express CD11b (22). While CD11d has received less attention than the other  $\beta_2$  integrins due to the absence of commercially available human

antibodies, Miyazaki and colleagues showed CD11d expression on monocyte-derived DCs and macrophages as well as most circulating monocytes (23). To complement the scarce available data, mRNA expression data for the CD11d subunit ITGAD were consulted. While Villani and colleagues (24) find monocytes to express highest levels of ITGAD mRNA, the Expression Atlas (25) reports highest expression in DCs, with ITGAD expression in monocytes remaining below detectable threshold. However, overall both RNAseq data sets show that CD11d mRNA expression is very low in monocytes, macrophages, and DCs. **Table 1**

**TABLE 1** |  $\beta_2$  integrin expression on dendritic cells (DCs), monocytes and macrophages—human and murine findings.

Cell type	CD11a/CD18 ( $\alpha_L/\beta_2$ )	CD11b/CD18 ( $\alpha_M/\beta_2$ )	CD11c/CD18 ( $\alpha_X/\beta_2$ )	CD11d/CD18 ( $\alpha_D/\beta_2$ )
DCs	<i>Human</i> : high levels of CD11a on monocyte-derived DCs (22, 27–29); plasmacytoid DCs (pDCs) also express CD11a (30); reduced CD11a/CD18 levels upon DC activation (31)	<i>Human</i> : CD11b present on monocyte-derived DCs (22, 27, 28); detected in cDCs, but not in pDCs (31–33); higher on cDC2 than cDC1s (33, 34); reduced CD11b/CD18 levels upon DC activation (31)	<i>Human</i> : pDCs lack CD11c (31); expressed on mature DCs (31); CD11c expression is higher on cDC2 than cDC1s (33, 34); monocyte-derived DCs also express CD11c (32); reduced CD11c/CD18 levels upon DC activation (31)	<i>Human</i> : expressed on monocyte-derived DCs (23), single-cell mRNA data suggests low gene expression in DCs (25)
	<i>Mouse</i> : expressed by cDCs, particularly the CD8 <sup>+</sup> subset, and by pDCs (35); also highly expressed by bone marrow-derived DCs	<i>Mouse</i> : expression of CD11b in mouse cDCs is subset-specific: higher on CD8 <sup>+</sup> than CD8 <sup>+</sup> splenic DCs (35); expressed in sub-populations of gut DCs (36); absent from pDCs (37); expressed by bone marrow-derived DCs (38)	<i>Mouse</i> : CD11c highly expressed on cDCs and typically used as a DC marker (38); expressed by pDCs (39) and bone marrow-derived DCs (40)	<i>Mouse</i> : no protein expression data available, RNA-seq data suggest medium ITGAD gene expression in murine DCs (25)
Monocytes	<i>Human</i> : expressed by circulating monocytes (21, 29, 41)	<i>Human</i> : highly expressed by circulating monocytes (21, 34, 41); differentially expressed on osteoclast precursors (42)	<i>Human</i> : expressed on circulating monocytes (21, 34) and classical, non-classical, and intermediate monocytes (31)	<i>Human</i> : expressed on majority of circulating monocytes, higher on CD16 <sup>+</sup> cells compared to CD16 <sup>+</sup> cells (23)
	<i>Mouse</i> : expressed by circulating monocytes (43)	<i>Mouse</i> : high expression of CD11b on murine monocytes (44)	<i>Mouse</i> : thought to be absent from most monocytes (45); though may be upregulated upon stimulation/maturation (44)	<i>Mouse</i> : lowly expressed by circulating monocytes, upregulated upon differentiation into macrophages (46), low ITGAD mRNA expression (25)
Macrophages	<i>Human</i> : expressed by monocyte-derived macrophages (21, 43); reduced expression on monocyte-derived macrophages compared to blood monocytes (21)	<i>Human</i> : expressed on monocyte-derived macrophages (47–49); expressed on alveolar macrophages, though at lower levels compared to blood monocytes (21)	<i>Human</i> : lowly expressed by monocyte-derived macrophages (21, 48–50)	<i>Human</i> : expressed on monocyte-derived macrophages <i>in vitro</i> (23)
	<i>Mouse</i> : expression dependent on tissue: present on pulmonary, but not on microglia, spleen or peritoneal macrophages (51)	<i>Mouse</i> : abundantly expressed by peritoneal macrophages (52, 53); highly expressed on dermal macrophages (54)	<i>Mouse</i> : expressed on alveolar macrophages (55); absent from bone marrow-derived macrophages and dermal macrophages (54)	<i>Mouse</i> : expressed by peritoneal macrophages (56)

provides the details of expression of all  $\beta_2$  integrin subunits in human and murine monocytes, macrophages, and DCs. Where available, expression analysis on DC subsets is given using the Guilliams nomenclature (26), which was recently confirmed and expanded by Villani and colleagues (24).

Animal studies have been instrumental in elucidating integrin function in monocytes, macrophages, and DCs.  $\beta_2$  integrins are highly conserved across species, with mice, rats, and rabbits most commonly used as models. Importantly  $\beta_2$  integrin-deficient mice are considered an appropriate model of the human condition leukocyte adhesion deficiency (LAD) where  $\beta_2$  integrin expression or function is lost (57). However, while  $\beta_2$  integrin structure is largely similar between species, cellular expression levels can vary significantly. A common example is CD11c, which in mice is predominantly expressed by conventional (cDCs) and plasmacytoid DCs (pDCs), although can also be expressed on lymphocyte subsets. In humans, on the other hand, CD11c is expressed not only on DCs but also monocytes, macrophages, granulocytes, and natural killer cells (19, 38). Animal and human studies therefore have to be compared with great care, and validation of concepts conceived in animal models in human cells remains a priority in elucidating the functions of  $\beta_2$  integrins.

## $\beta_2$ INTEGRINS AS REGULATORS OF IMMUNE FUNCTION

### Evidence for $\beta_2$ Integrin Contribution to Immune Regulation

There is mounting evidence that puts  $\beta_2$  integrins at the center of the balance between immune priming and tolerance. Integrin-deficient humans and mouse models show that  $\beta_2$  integrins are important negative regulators of the immune system. LADs are genetic human disorders caused by the reduction or complete absence of  $\beta_2$ -integrins (LAD-I) (58) or by mutations in the integrin-activating protein kindlin-3 (LAD-III) (59). These disorders are characterized by profound impairment of leukocyte recruitment to peripheral sites of infection. Patients with LAD suffer from increased susceptibility to infection and impaired inflammatory responses (60), resulting in markedly reduced lifespan if no therapeutic measures are taken. Paradoxically LAD patients also suffer from chronic inflammatory diseases. Examples of conditions prevalent in LAD patients include intestinal colitis (61) and periodontitis (62) suggest that  $\beta_2$  integrins have an important role in suppressing inflammation and promoting



immune tolerance. Supporting this, the presence of functional  $\beta_2$  integrins improved symptoms in a model of skin inflammation by restricting DC-mediated T cell activation (63).

LAD pathology can be replicated in  $\beta_2$  integrin knockout (KO) mouse models, underlining the importance of  $\beta_2$  integrins for immune cell recruitment in both humans and murine models and the similarities between the species. From studies in KO mice and LAD patients, we know  $\beta_2$  integrins are essential in mediating T cell recruitment to lymph nodes and leukocyte, particularly neutrophil and T cell, recruitment to sites of inflammation. Here, we will further explore the roles of these integrins in monocytes, macrophages, and DCs.

## $\beta_2$ Integrins Regulate Recruitment and Migration of Mononuclear Phagocytes

Evidence suggests that leukocyte recruitment to tissues is dependent on  $\beta_2$  integrins, because of the requirement for these adhesion molecules in the firm adhesion to the endothelial layer under shear flow conditions and for subsequent transendothelial migration (64). However, leukocyte migration *within* tissues is thought to occur independently of  $\beta_2$  integrins, as cells use an actin-dependent flowing and squeezing mechanism of movement in three-dimensional environments (64).

Geissmann and colleagues showed that the adhesion of patrolling murine monocytes to blood vessel walls is significantly decreased when CD11a is blocked (45). Similarly, chemotactic migration of human monocytes *in vitro* is inhibited when CD18 function is blocked (65). However, murine monocyte recruitment to sites of inflammation was found to occur independently of CD11a and CD11b (66), suggesting that  $\beta_2$  integrins are primarily involved in the homeostatic migration of monocytes and that their role is redundant during inflammation. On the other hand, increased expression levels of CD11d on macrophages mediates their retention at inflammatory sites in mice (56).

The role of  $\beta_2$  integrins in DC and macrophage recruitment to secondary lymphoid organs and tissues seems to be dependent on the inflammatory state of the body. Bone marrow-derived DCs (BMDCs) from mice where all integrins, including  $\beta_2$ , are knocked out, migrated from the site of injection (ear) to the draining lymph node in similar numbers to their wild-type counterparts when activated with lipopolysaccharide (LPS). This suggests that DC migration during inflammation is not dependent on integrins. However, under steady-state conditions, the absence of functional  $\beta_2$  integrins from murine BMDCs (using signaling-deficient  $\beta_2$  integrin knock-in BMDCs) was found to increase migration from tissue (footpad) to draining lymph node, leading to the hypothesis that  $\beta_2$  integrins function to restrict migration in the steady-state by anchoring DCs in the tissue site. As a consequence of increased DC migration to the draining lymph node, the same study showed an increase in Th1 cytokine production (67), further supporting a negative regulatory role for  $\beta_2$  integrins on DCs. In addition, a murine model of skin inflammation also showed an increase in migratory DCs in the draining lymph node of  $\beta_2$  integrin signaling-deficient mice, as well as at the site of inflammation, though whether this was dependent on the inflammation or not was not determined (63). Overall, the cellular environment seems to determine the requirement for

functional  $\beta_2$  integrins in the migration of both monocytes and DCs *in vivo*: integrins play a role in monocyte recruitment and DC migration under steady-state conditions, but are dispensable during inflammation.

## $\beta_2$ Integrins Regulating DC–T Cell Interactions

In addition to their roles in leukocyte recruitment and migration,  $\beta_2$  integrins are also important mediators of cellular interactions. Functional  $\beta_2$  integrins are important in the formation of the immunological synapse between antigen-presenting cells (APCs) and T cells. The context and dynamics of this interaction determine whether T cells become activated or tolerized.  $\beta_2$  integrins, and their ligand, ICAM-1, are expressed by both the T cell and the APC and are vital in immune synapse formation. Importantly, it is becoming increasingly clear that  $\beta_2$  integrins expressed by the APC and T cell have opposing functions in the immune synapse, resulting in differential outcomes for the T cell response.

On the T cell side, CD11a clusters in the peripheral supramolecular activation cluster (P-SMAC) and binds to ICAM-1 on the APC (68). This molecular interaction stabilizes the connection made between T cell receptor and peptide:MHC on the APC in the central SMAC (16, 69), thereby enhancing TCR signal transduction (70). While T cell CD11a therefore has a largely pro-inflammatory effect, enhancing T cell activation, proliferation, and differentiation, a role for T cell integrins in regulation of activation, for example, in different T cell subsets, is not ruled out.

On the APC side of the immunological synapse,  $\beta_2$  integrins have also been shown to be involved, likely binding to ICAM-1 on the T cell. Importantly, the integrins on the APC regulate the outcome of the T cell response. For example, in murine models, active CD11b on DC surfaces inhibits the DC–T cell interaction (71). The reduced antigen-presenting capabilities of murine bone marrow-derived macrophages compared to BMDCs were therefore proposed to be due to their comparably larger surface expression of activated CD11b (71, 72). This suppressive role for DC CD11b has also been shown in human cells. When CD11b on human monocyte-derived DCs binds its ligand ICAM-1, both CD86 expression on DCs and DC-induced T cell proliferation were reduced (73). Interestingly, ligation of CD11b/CD18 decreases the ability of murine BMDCs to stimulate T cells and elicit a downstream response (74), CD11b/CD18 interactions can suppress Th17 cell differentiation (75), suggesting a strong role for this specific  $\beta_2$  integrin in immune regulation. This suggests that the activated conformation of CD11b/CD18 is extensively involved in regulating the immune system and has strong negative and positive regulatory functions depending on cell type they are expressed on.

Furthermore, the expression of activated  $\beta_2$  integrins on murine DC surfaces significantly reduces T cell activation (71) and further studies actually demonstrated an inverse relationship between forced activation of murine BMDC CD11a and T cell activation (72), suggesting a directly limiting effect of active  $\beta_2$  integrins on T cell activation by APCs.

Overall, the role of integrins as adhesion molecules carefully mediating and regulating cellular interactions is not to be underestimated for mounting an effective immune response.

## $\beta_2$ Integrins Regulate Immune Cell Signaling

In addition to their roles in leukocyte recruitment and interactions, several studies show that integrin outside-in signaling following ligand binding can directly affect cell function. Chinese Hamster Ovarian cells transfected with CD11c acquire the ability to bind both LPS and Gram-negative bacteria, as well as the ability to initiate downstream activation signals (76). In contrast to their anti-inflammatory roles on DCs, CD11b or CD11c receptor occupation on the surface of human monocytes stimulates cell-specific pro-inflammatory pathways (77), such as secretion of IL-8, MIP1 $\alpha$ , and MIP1 $\beta$ .

Generally, the interplay between TLR4- and  $\beta_2$  integrin-mediated signaling is controversial. On the one hand, it has been shown that CD11b positively regulates TLR4 signaling (78), especially in murine BMDCs. Several studies report  $\beta_2$  integrins act in synergy with LPS (79–81), therefore suggesting a potential pro-inflammatory role for CD11b. By contrast, other studies report that  $\beta_2$  integrins negatively affect TLR signaling. Complete absence of  $\beta_2$  integrins in mice (CD18 KO) was shown to result in a strong increase of TLR signaling (82) and the absence of CD11b specifically from murine macrophages causes exacerbated TLR-mediated inflammatory responses, resulting in increased susceptibility to endotoxin shock and *Escherichia coli* sepsis (83). Mechanistically, CD11b signaling has been shown to induce degradation of the key TLR signaling components, MyD88 and TRIF, directly dampening TLR responses in macrophages (83). Moreover, activation of CD11b on human inflammatory arthritis synovial macrophages *via* binding to its ligand ICAM was shown to indirectly inhibit TLR signaling (84) by inducing expression of IL-10 and the inhibitory factors SOCS3, ABIN-3, and A20. Integrins furthermore restrict TLR signaling on both murine macrophages and DCs (63). The role of  $\beta_2$  integrins in modulating TLR signaling is, therefore, complex, although one could tentatively propose that CD11b specifically seems to have opposing TLR4-mediated roles in inflammation, depending on the APC surface it is expressed on. However, while this could hold true for TLR4 signaling, this might not be the case for all TLRs. CD11b deficiency in murine BMDCs, while negatively affecting TLR4-mediated pathways, actually leads to an increase

in DC cross-priming of cytotoxic T cells, a process mediated by the microRNA-146a (85).  $\beta_2$  integrin regulation of TLR-mediated responses therefore remains incompletely understood, with future studies hopefully elucidating the complex and intricate nature of these receptor interactions.

A variety of studies available suggest a significant immunoregulatory role for  $\beta_2$  integrins, not only by their mediation of adhesive and migratory processes, but also by immunological signaling. However, other studies suggest that, given the right cellular environment or cell type,  $\beta_2$  integrins can also have a strong pro-inflammatory effect (see **Table 2** for comparison). When considering these opposing functions of integrins, it seems likely that even slight disturbances in integrin expression, signaling or activation could result in significant immunological effects, thus potentially contributing to a variety of autoimmune, inflammatory, and infectious conditions.

## $\beta_2$ INTEGRINS IN INFLAMMATION, INFECTION, AND AUTOIMMUNITY

Evidence for the role of  $\beta_2$  integrins in contributing to the development and progression of inflammatory and autoimmune conditions is accumulating. Considering that  $\beta_2$  integrin signaling can have opposing functions depending on subunit pairing and the immune cell type it is expressed on, it is not surprising that these receptors play important roles in both *contributing to* as well as *negatively regulating* inflammatory processes.

Human genetic studies point to a role of  $\beta_2$  integrins in inflammation and autoimmunity. A polymorphism of *ITGAM*, the CD11b subunit, increases the risk for the autoimmune disease systemic lupus erythematosus (87) (SLE), which shares genetic risk factors with rheumatoid arthritis (RA) (88). Disease risk for inflammatory bowel disease, similarly characterized by dysregulation of immune function specifically in the intestine, increases with amplified expression of alleles for both *ITGAL*, encoding CD11a, and the  $\beta_2$  integrin ligand *ICAM1* (89). Gene expression of CD11d in humans and mice was found to be increased in white adipose tissue in obesity, a condition characterized by an increase in systemic inflammation (90). Furthermore, CD11d activation led to increased IL-1 $\beta$  expression (23), which when overproduced

**TABLE 2** | Summary of the roles for  $\beta_2$  integrins in monocytes, macrophages, and dendritic cells (DCs).

Cell type	Recruitment and migration	Interactions with T cells	Signaling
Monocytes	$\beta_2$ integrins mediate recruitment of monocytes under homeostatic conditions (45, 65), but dispensable for recruitment during inflammation (66)	Yet to be determined	Yet to be determined
	<i>Pro-inflammatory</i>	<i>Unknown</i>	<i>Unknown</i>
Macrophages	$\beta_2$ integrins reported to mediate macrophage retention at inflammatory sites (56, 86)	Yet to be determined	$\beta_2$ integrin signaling dampens macrophage responses to Toll-like receptor (TLR) stimulation (82, 83)
	<i>Pro-inflammatory</i>	<i>Unknown</i>	<i>Regulatory</i>
DCs	Under homeostatic conditions $\beta_2$ integrins restrict DC migration from tissue to lymph nodes (67); Migration from tissue site to draining lymph nodes during inflammation occurs independently of integrins (64)	DC integrins contribute to contact formation with T cells—this role inhibits full T cell activation (71, 72, 74)	$\beta_2$ integrin signaling functions to restrict DC activation both in response to TLR stimulation and under homeostatic conditions (67)
	<i>Regulatory</i>	<i>Regulatory</i>	<i>Regulatory</i>

can contribute to a variety of autoinflammatory conditions (91). While dysregulation of  $\beta_2$  integrin signaling seems likely to be involved in a variety of autoimmune diseases and inflammatory conditions, exact mechanisms are still unclear, and further investigation of both signaling pathways and genetic basis will be needed to fully elucidate their complex roles.

Recent studies have focused on  $\beta_2$  integrin involvement in RA, which serves as an excellent example of the opposing roles  $\beta_2$  integrins can take in disease. Expression of CD11a is increased in inflamed synovial tissue, where it is hypothesized to contribute to cell activation and on-going joint destruction (92, 93), but not in peripheral blood of RA patients. However, as CD11a is also involved in facilitating immune cell migration to sites of inflammation, clear-cut cause and effect of the presence of activated  $\beta_2$  integrins in the synovium is difficult to establish. Blocking all  $\beta_2$  integrins reduced inflammation in a rabbit RA model (94), while absence of CD11a led to complete resistance to disease induction in a KB  $\times$  N serum transfer mouse model of arthritis (95). Furthermore, both a small molecule antagonist against CD11a and a CD11a-monoclonal antibody (mAb) proved to be similarly successful in reducing both inflammatory-mediated bone destruction and cytokine mRNA levels within the murine joint (96, 97). Mice with mutations in the  $\beta_2$  integrin ligand ICAM-1 also show reduced susceptibility to the collagen-induced arthritis (CIA) model (98). Clearly, CD11a-ICAM-1 interactions are essential for leukocyte recruitment to the inflamed joint.

However, evidence is emerging that other  $\beta_2$  integrins may function to control inflammation in arthritis. CD11b KO mice, for example, show exacerbated joint pathology in the KB  $\times$  N serum transfer model of arthritis, underlining the starkly opposite roles different  $\beta_2$  integrins can play (95). A recent study replicated these results in a CIA model and, furthermore, showed that exacerbated joint pathology resulted from elevated IL-6 levels and an increase in Th17 cell priming, which could be rescued by introducing a CD11b-expressing DC cell line (99). On the other hand, blocking CD11b immediately before onset of disease significantly reduced disease burden in two different models of arthritis (CIA and a DBA/1 to severe combined immunodeficiency transfer model of arthritis) (100), suggesting that the role of CD11b in inflammatory arthritis may differ depending on the cell type involved and the disease stage.

When considering the importance, as well as the obvious complexity, of  $\beta_2$  integrin function in autoimmune diseases such as RA, therapeutically targeting  $\beta_2$  integrins will have to be carefully balanced but also holds great promise to offer novel treatment options.

## APPLICABILITY OF INTEGRIN-TARGETING THERAPIES

Modulating integrin function to improve mal-adaptation or excessive activation of the immune system is of great interest in a variety of autoimmune and inflammatory conditions. However, achieving efficacy without immunocompromising side effects might prove challenging. Here, we discuss the progress and failures in developing integrin-targeted therapies and speculate on the routes forward for success.

To date, targeting integrins therapeutically has had mixed success in the clinic. The only mAb targeting  $\beta_2$  integrins, Efalizumab, which targets CD11a, was originally developed as a treatment for psoriasis (101). However, several patients presented with the potentially fatal disease progressive multifocal leukoencephalopathy (PML), caused by reactivation of the JC virus, which results in a white matter disorder of the brain (102). Although the mechanism of PML development in Efalizumab-treated patients was not investigated, we speculate that viral reactivation was likely either due to the loss of immune cell recruitment to the brain to control the virus (103) or due to the mAb itself crossing the blood-brain barrier (104). Due to the occurrence of PML, Efalizumab was withdrawn from European and American markets due to its associated safety issues in 2009.

Although targeting  $\beta_2$  integrins has so far failed in the clinic, targeting other integrins for the treatment of colitis and Crohn's disease has proved successful. The mAb against the  $\alpha_4$  integrin, Natalizumab, was developed for the treatment of multiple sclerosis and Crohn's disease (105, 106). This mAb binds to  $\alpha_4\beta_1$  and  $\alpha_4\beta_7$ . However, PML also occurs in some Natalizumab-treated patients (integrin  $\alpha_4\beta_1$  is also involved in leukocyte recruitment to the brain) and so is no longer used widely (107). More recently, a specific  $\alpha_4\beta_7$  targeting mAb Vedolizumab has shown success in safety efficacy in Crohn's disease and ulcerative colitis. This success story underlines the potential of targeting integrins for therapeutic purposes.

In order to realize the potential of targeting  $\beta_2$  integrins therapeutically, it will be necessary to improve the strategy. As indicated by the success of Vedolizumab over Natalizumab, one way to do this is to target the right integrin subunit(s) in order to reduce the likelihood of side effects. Targeting CD11a, in the form of Efalizumab, proved unsuccessful in the clinic. As CD11a is expressed by almost all leukocytes, has vital roles in leukocyte recruitment and has immunoregulatory effects in mononuclear phagocytes, the resulting serious side effects from targeting this molecule therapeutically are, perhaps, not surprising. Targeting other CD11 subunits might be a more effective strategy. For example, CD11b, CD11c, and CD11d have a more restricted pattern of expression in leukocytes (predominantly on monocytes, macrophages, and DCs), which may make these molecules more suitable targets. Importantly, it is vital that we consider the pro- and anti-inflammatory functions of  $\beta_2$  integrin subunits and design drugs to target them appropriately. CD11b, for example, has clear regulatory roles in macrophages and DCs, meaning that we could potentially exploit this immunosuppressive pathway by activating, rather than blocking, this integrin subunit. Such a strategy may have less risk of serious side effects. It is, therefore, essential that we fully understand the specific functions of individual integrin subunits in different leukocyte populations in order to target  $\beta_2$  integrin subunits effectively in the clinic.

Another option to explore is blocking not the  $\beta_2$  integrin itself, but the ligand of interest. Targeting the CD11a and CD11b ligand, ICAM-1, has shown beneficial results especially in early RA (108), although immunogenicity of the mAb in question restricts clinical use (109) and problems caused by impaired leukocyte recruitment prevail.



Further potential difficulties in developing integrin-targeting therapy include the close signaling relationships that exist in some integrins, potentially leading to complex downstream effects mediated even by an activating mAb highly specific for a  $\beta_2$  integrin (110). Carefully elucidating downstream signaling pathways and further increasing drug specificity is therefore essential to bring more integrin therapeutics into the clinic.

Innovative avenues to explore include computationally designed integrin proteins with constitutively activated or inactivated subunits, which could find applications in both pharmacological testing and therapy (111). Furthermore, developing small molecular drugs targeting  $\beta_2$  integrins viable for oral use remains a priority, as it could offer an alternative way to yield the same beneficial results without the dangerous side effects of mAbs. An example is the small molecule CD11b agonist, Leukadherin-1, which previous studies found to reduce monocyte-mediated TNF-release by mimicking natural ligand binding. When NK cells and monocytes were pre-treated with Leukadherin-1, innate inflammatory signaling in human *ex vivo* studies was suppressed (112). While the study noted some caveats, for example, the differences of CD11b function on different cell types (78), the drug is still being explored for the treatment of SLE. Another small molecule currently in development is the CD11a antagonist BMS-587101, which acts by reducing CD11a-mediated adhesion and to a lesser effect T cell proliferation. It significantly improved both murine models of lung inflammation and transplant viability (113).

Continuous effort to increase drug specificity and further understand their complex delicate signaling networks will be needed to bring  $\beta_2$  integrin-targeting drugs into the clinic. But while the use of integrin-targeting drugs has been contentious in the past, their potential in treating a wide variety of immune diseases is enormous and should not be neglected.

## CONCLUSION

This review explored the opposing nature of  $\beta_2$  integrin pro- and anti-inflammatory functions in three main immune functions,

making them prime candidates to be both important mediators and regulators of the immune system. The first is migration, which allows for targeted immune cell recruitment to sites of infection and tissue damage. The second is adhesion, not only preceding immune cell extravasation at sites of inflammation, but also an important factor in initiating the adaptive immune response by facilitating cellular interactions. Finally, immune cell signaling, which allows for fine-tuned cooperation between a wide variety of immune cells. Considering the fact that  $\beta_2$  integrins play a complex role in three important areas of the immune system and their differential expression on monocytes, macrophages and DCs, it becomes clear that the variety of studies presented in this review is by no means exhaustive. The common message is evident:  $\beta_2$  integrins are involved in complex immunoregulatory signaling pathways. However, in addition to their well-established pro-inflammatory roles in recruitment and activation,  $\beta_2$  integrins also have essential immunoregulatory functions. Dysregulated integrin signaling, expression and surface activation is therefore likely to contribute to a variety of inflammatory and autoimmune conditions. Elucidating the function of  $\beta_2$  integrins further therefore promises to provide novel therapeutic targets for various disorders, RA being just one example.

## AUTHOR CONTRIBUTIONS

CH and VM designed the structure of the review. LS wrote the first draft. CH and VM revised the manuscript. LS composed the figures. All authors have seen and agreed on the finally submitted version of the manuscript.

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## REFERENCES

1. Mould AP, Humphries MJ. Regulation of integrin function through conformational complexity: not simply a knee-jerk reaction? *Curr Opin Cell Biol* (2004) 16(5):544–51. doi:10.1016/j.ccb.2004.07.003
2. Adair BD, Xiong JP, Maddock C, Goodman SL, Arnaout MA, Yeager M. Three-dimensional EM structure of the ectodomain of integrin  $\alpha V\beta 3$  in a complex with fibronectin. *J Cell Biol* (2005) 168(7):1109–18. doi:10.1083/jcb.200410068
3. Byron A, Humphries JD, Askari JA, Craig SE, Mould AP, Humphries MJ. Anti-integrin monoclonal antibodies. *J Cell Sci* (2009) 122(22):4009–11. doi:10.1242/jcs.056770
4. Stewart M, Hogg N. Regulation of leukocyte integrin function: affinity vs. avidity. *J Cell Biochem* (1996) 61(4):554–61. doi:10.1002/(SICI)1097-4644(19960616)61:4<554::AID-JCB8>3.0.CO;2-N
5. Hogg N, Patzak I, Willenbrock F. The insider's guide to leukocyte integrin signalling and function. *Nat Rev Immunol* (2011) 6:416–26. doi:10.1038/nri2986
6. Harburger DS, Calderwood DA. Integrin signalling at a glance. *J Cell Sci* (2009) 122(2):159–63. doi:10.1242/jcs.018093
7. Tadokoro S, Shattil SJ, Eto K, Tai V, Liddington RC, de Pereda JM, et al. Talin binding to integrin  $\beta$  tails: a final common step in integrin activation. *Science* (2003) 302(5642):103–6. doi:10.1126/science.1086652
8. Moser M, Nieswandt B, Ussar S, Pozgajova M, Fässler R. Kindlin-3 is essential for integrin activation and platelet aggregation. *Nat Med* (2008) 14(3):325–30. doi:10.1038/nm1722
9. Moser M, Bauer M, Schmid S, Ruppert R, Schmidt S, Sixt M, et al. Kindlin-3 is required for  $\beta_2$  integrin-mediated leukocyte adhesion to endothelial cells. *Nat Med* (2009) 15(3):300–5. doi:10.1038/nm.1921
10. Constantin G, Majeed M, Giagulli C, Piccio L, Kim JY, Butcher EC, et al. Chemokines trigger immediate  $\beta_2$  integrin affinity and mobility changes: differential regulation and roles in lymphocyte arrest under flow. *Immunity* (2000) 13(6):759–69. doi:10.1016/S1074-7613(00)00074-1
11. Chung KJ, Mitroulis I, Wiessner JR, Zheng YY, Siegert G, Sperandio M, et al. A novel pathway of rapid TLR-triggered activation of integrin-dependent leukocyte adhesion that requires Rap1 GTPase. *Mol Biol Cell* (2014) 25(19):2948–55. doi:10.1091/mbc.E14-04-0867
12. Bazzoni G, Hemler ME. Are changes in integrin affinity and conformation overemphasized? *Trends Biochem Sci* (1998) 23(1):30–4. doi:10.1016/S0968-0004(97)01141-9

13. Carman CV, Springer TA. Integrin avidity regulation: are changes in affinity and conformation underemphasized? *Curr Opin Cell Biol* (2003) 15(5):547–56. doi:10.1016/j.ceb.2003.08.003
14. Iwamoto DV, Calderwood DA. Regulation of integrin-mediated adhesions. *Curr Opin Cell Biol* (2015) 36:41–7. doi:10.1016/j.ceb.2015.06.009
15. von Andrian UH, Chambers JD, McEvoy LM, Bargatze RF, Arfors KE, Butcher EC. Two-step model of leukocyte-endothelial cell interaction in inflammation: distinct roles for LECAM-1 and the leukocyte beta 2 integrins in vivo. *Proc Natl Acad Sci U S A* (1991) 88(17):7538–42. doi:10.1073/pnas.88.17.7538
16. Monks CRF, Freiberg BA, Kupfer H, Sciaky N, Kupfer A. Three-dimensional segregation of supramolecular activation clusters in T cells. *Nature* (1998) 395(6697):82–6. doi:10.1038/25764
17. Podolnikova NP, Podolnikov AV, Haas TA, Lishko VK, Ugarova TP. Ligand recognition specificity of leukocyte integrin  $\alpha\text{M}\beta_2$  (Mac-1, CD11b/CD18) and its functional consequences. *Biochemistry* (2015) 54(6):1408–20. doi:10.1021/bi5013782
18. Yakubenko VP, Yadav SP, Ugarova TP. Integrin  $\alpha\text{D}\beta_2$ , an adhesion receptor up-regulated on macrophage foam cells, exhibits multiligand-binding properties. *Blood* (2006) 107(4):1643–50. doi:10.1182/blood-2005-06-2509
19. Sándor N, Lukácsi S, Ungai-Salánki R, Orgován N, Szabó B, Horváth R, et al. CD11c/CD18 dominates adhesion of human monocytes, macrophages and dendritic cells over CD11b/CD18. *PLoS One* (2016) 11(9):e0163120. doi:10.1371/journal.pone.0163120
20. Erdei A, Lukácsi S, Mácsik-Valent B, Nagy-Baló Z, Kurucz I, Bajtay Z. Non-identical twins: different faces of CR3 and CR4 in myeloid and lymphoid cells of mice and men. *Semin Cell Dev Biol* (2017). doi:10.1016/j.semcdb.2017.11.025
21. Prieto J, Eklund A, Patarroyo M. Regulated expression of integrins and other adhesion molecules during differentiation of monocytes into macrophages. *Cell Immunol* (1994) 156(1):191–211. doi:10.1006/cimm.1994.1164
22. Freudenthal PS, Steinman RM. The distinct surface of human blood dendritic cells, as observed after an improved isolation method. *Proc Natl Acad Sci U S A* (1990) 87(19):7698–702. doi:10.1073/pnas.87.19.7698
23. Miyazaki Y, Vieira-de-Abreu A, Harris ES, Shah AM, Weyrich AS, Castro-Faria-Neto HC, et al. Integrin  $\alpha\text{D}\beta_2$  (CD11d/CD18) is expressed by human circulating and tissue myeloid leukocytes and mediates inflammatory signaling. *PLoS One* (2014) 9(11):e112770. doi:10.1371/journal.pone.0112770
24. Villani AC, Satija R, Reynolds G, Sarkizova S, Shekhar K, Fletcher J, et al. Single-cell RNA-seq reveals new types of human blood dendritic cells, monocytes, and progenitors. *Science* (2017) 356(6335):eaah4573. doi:10.1126/science.aah4573
25. EMBL-EBI Expression Atlas Development Team, github.com/gxa/atlas/graphs/contributors. Expression atlas. *EMBL-EBI Gene Expression Atlas*. European Molecular Biology Laboratory - European Bioinformatics Institute (2017). Available from: <http://www.ebi.ac.uk/gxa/home>
26. Guillems M, Ginhoux F, Jakubczik C, Naik SH, Onai N, Schraml BU, et al. Dendritic cells, monocytes and macrophages: a unified nomenclature based on ontogeny. *Nat Rev Immunol* (2014) 14(8):571–8. doi:10.1038/nri3712
27. Sallusto F, Lanzavecchia A. Efficient presentation of soluble antigen by cultured human dendritic cells is maintained by granulocyte/macrophage colony-stimulating factor plus interleukin 4 and downregulated by tumor necrosis factor. *J Exp Med* (1994) 179(4):1109–18. doi:10.1084/jem.179.4.1109
28. Geijtenbeek TB, Torensma R, van Vliet SJ, van Duijnhoven GC, Adema GJ, van Kooyk Y, et al. Identification of DC-SIGN, a novel dendritic cell-specific ICAM-3 receptor that supports primary immune responses. *Cell* (2000) 100(5):575–85. doi:10.1016/S0092-8674(00)80693-5
29. Ammon C, Meyer SP, Schwarzfischer L, Krause SW, Andreesen R, Kreutz M. Comparative analysis of integrin expression on monocyte-derived macrophages and monocyte-derived dendritic cells. *Immunology* (2000) 100(3):364–9. doi:10.1046/j.1365-2567.2000.00056.x
30. de la Rosa G, Longo N, Rodríguez-Fernández JL, Puig-Kroger A, Pineda A, Corbí AL, et al. Migration of human blood dendritic cells across endothelial cell monolayers: adhesion molecules and chemokines involved in subset-specific transmigration. *J Leukoc Biol* (2003) 73(5):639–49. doi:10.1189/jlb.1002516
31. Rieckmann JC, Geiger R, Hornburg D, Wolf T, Kveler K, Jarrossay D, et al. Social network architecture of human immune cells unveiled by quantitative proteomics. *Nat Immunol* (2017) 18(5):583–93. doi:10.1038/ni.3693
32. Li K, Fazekasova H, Wang N, Sagoo P, Peng Q, Khamri W, et al. Expression of complement components, receptors and regulators by human dendritic cells. *Mol Immunol* (2011) 48(9):1121–7. doi:10.1016/j.molimm.2011.02.003
33. Haniffa M, Shin A, Bigley V, McGovern N, Teo P, See P, et al. Human tissues contain CD141 hi cross-presenting dendritic cells with functional homology to mouse CD103+ nonlymphoid dendritic cells. *Immunity* (2012) 37(1):60–73. doi:10.1016/j.immuni.2012.04.012
34. Robbins SH, Walzer T, Dembélé D, Thibault C, Defays A, Bessou G, et al. Novel insights into the relationships between dendritic cell subsets in human and mouse revealed by genome-wide expression profiling. *Genome Biol* (2008) 9(1):R17. doi:10.1186/gb-2008-9-1-r17
35. Segura E, Guérin C, Hogg N, Amigorena S, Théry C. CD8+ dendritic cells use LFA-1 to capture MHC-peptide complexes from exosomes in vivo. *J Immunol* (2007) 179(3):1489–96. doi:10.4049/jimmunol.179.3.1489
36. Cerovic V, Bain CC, Mowat AM, Milling SW. Intestinal macrophages and dendritic cells: what's the difference? *Trends Immunol* (2014) 35(6):270–7. doi:10.1016/j.it.2014.04.003
37. Nakano H, Yanagita M, Gunn MD. CD11c+ B220+ Gr-1+ cells in mouse lymph nodes and spleen display characteristics of plasmacytoid dendritic cells. *J Exp Med* (2001) 194(8):1171–8. doi:10.1084/jem.194.8.1171
38. Shortman K, Liu YJ. Mouse and human dendritic cell subtypes. *Nat Rev Immunol* (2002) 2(3):151–61. doi:10.1038/nri746
39. Miller JC, Brown BD, Shay T, Gautier EL, Jojic V, Cohain A, et al. Deciphering the transcriptional network of the dendritic cell lineage. *Nat Immunol* (2012) 13(9):888–99. doi:10.1038/ni.2370
40. Helft J, Böttcher J, Chakravarty P, Zelenay S, Huotari J, Schraml BU, et al. GM-CSF mouse bone marrow cultures comprise a heterogeneous population of CD11c+ MHCII+ macrophages and dendritic cells. *Immunity* (2015) 42(6):1197–211. doi:10.1016/j.immuni.2015.05.018
41. Bohuslav J, Horejsi V, Hansmann C, Stöckl J, Weidle UH, Majdic O, et al. Urokinase plasminogen activator receptor, beta 2-integrins, and Src-kinases within a single receptor complex of human monocytes. *J Exp Med* (1995) 181(4):1381–90. doi:10.1084/jem.181.4.1381
42. Sprangers S, Schoenmaker T, Cao Y, Everts V, de Vries TJ. Integrin  $\alpha\text{M}\beta_2$  is differently expressed by subsets of human osteoclast precursors and mediates adhesion of classical monocytes to bone. *Exp Cell Res* (2017) 350(1):161–8. doi:10.1016/j.yexcr.2016.11.018
43. Auffray C, Fogg D, Garfa M, Elain G, Join-Lambert O, Kayal S, et al. Monitoring of blood vessels and tissues by a population of monocytes with patrolling behavior. *Science* (2007) 317(5838):666–70. doi:10.1126/science.1142883
44. Sunderkötter C, Nikolic T, Dillon MJ, Van Rooijen N, Stehling M, Drevets DA, et al. Subpopulations of mouse blood monocytes differ in maturation stage and inflammatory response. *J Immunol* (2004) 172(7):4410–7. doi:10.4049/jimmunol.172.7.4410
45. Geissmann F, Jung S, Littman DR. Blood monocytes consist of two principal subsets with distinct migratory properties. *Immunity* (2003) 19(1):71–82. doi:10.1016/S1074-7613(03)00174-2
46. Noti JD. Expression of the myeloid-specific leukocyte integrin gene CD11d during macrophage foam cell differentiation and exposure to lipoproteins. *Int J Mol Med* (2002) 10:721–8. doi:10.3892/ijmm.10.6.721
47. Wright SD, Jong MT. Adhesion-promoting receptors on human macrophages recognize *Escherichia coli* by binding to lipopolysaccharide. *J Exp Med* (1986) 164(6):1876–88. doi:10.1084/jem.164.6.1876
48. Mevorach D, Mascarenhas JO, Gershov D, Elkon KB. Complement-dependent clearance of apoptotic cells by human macrophages. *J Exp Med* (1998) 188(12):2313–20. doi:10.1084/jem.188.12.2313
49. Schlesinger LS, Horwitz MA. Phagocytosis of *Mycobacterium leprae* by human monocyte-derived macrophages is mediated by complement receptors CR1 (CD35), CR3 (CD11b/CD18), and CR4 (CD11c/CD18) and IFN-gamma activation inhibits complement receptor function and phagocytosis of this bacterium. *J Immunol* (1991) 147(6):1983–94.
50. Perrone LA, Plowden JK, García-Sastre A, Katz JM, Tumpey TM. H5N1 and 1918 pandemic influenza virus infection results in early and excessive infiltration of macrophages and neutrophils in the lungs of mice. *PLoS Pathog* (2008) 4(8):e1000115. doi:10.1371/journal.ppat.1000115
51. Gautier EL, Shay T, Miller J, Greter M, Jakubczik C, Ivanov S, et al. Gene-expression profiles and transcriptional regulatory pathways that underlie

- the identity and diversity of mouse tissue macrophages. *Nat Immunol* (2012) 13(11):1118–28. doi:10.1038/ni.2419
52. Beller DI, Springer TA, Schreiber RD. Anti-Mac-1 selectively inhibits the mouse and human type three complement receptor. *J Exp Med* (1982) 156(4):1000–9. doi:10.1084/jem.156.4.1000
  53. Drevets DA, Leenen PJ, Campbell PA. Complement receptor type 3 (CD11b/CD18) involvement is essential for killing of *Listeria monocytogenes* by mouse macrophages. *J Immunol* (1993) 151(10):5431–9.
  54. Tamoutounour S, Williams M, Montanana Sanchis F, Liu H, Terhorst D, Malosse C, et al. Origins and functional specialization of macrophages and of conventional and monocyte-derived dendritic cells in mouse skin. *Immunity* (2013) 39(5):925–38. doi:10.1016/j.immuni.2013.10.004
  55. Vermaelen K, Pauwels R. Accurate and simple discrimination of mouse pulmonary dendritic cell and macrophage populations by flow cytometry: methodology and new insights. *Cytometry A* (2004) 61(2):170–7. doi:10.1002/cyto.a.20064
  56. Yakubenko VP, Belevych N, Mishchuk D, Schurin A, Lam SC, Ugarova TP. The role of integrin  $\alpha$ D  $\beta$  2 (CD11d/CD18) in monocyte/macrophage migration. *Exp Cell Res* (2008) 314(14):2569–78. doi:10.1016/j.yexcr.2008.05.016
  57. Scharffetter-Kochanek K, Lu H, Norman K, van Nood N, Munoz F, Grabbe S, et al. Spontaneous skin ulceration and defective T cell function in CD18 null mice. *J Exp Med* (1998) 188(1):119–31. doi:10.1084/jem.188.1.119
  58. Harris ES, Weyrich AS, Zimmerman GA. Lessons from rare maladies: leukocyte adhesion deficiency syndromes. *Curr Opin Hematol* (2013) 20(1):16. doi:10.1097/MOH.0b013e32835a0091
  59. Robert P, Canault M, Farnarier C, Nurden A, Grosdidier C, Barlogis V, et al. A novel leukocyte adhesion deficiency III variant: kindlin-3 deficiency results in integrin- and nonintegrin-related defects in different steps of leukocyte adhesion. *J Immunol* (2011) 186(9):5273–83. doi:10.4049/jimmunol.1003141
  60. Kishimoto TK, Hollander N, Roberts TM, Anderson DC, Springer TA. Heterogeneous mutations in the  $\beta$  subunit common to the LFA-1, Mac-1, and p150, 95 glycoproteins cause leukocyte adhesion deficiency. *Cell* (1987) 50(2):193–202. doi:10.1016/0092-8674(87)90215-7
  61. D'agata ID, Paradis K, Chad Z, Bonny Y, Seidman E. Leucocyte adhesion deficiency presenting as a chronic ileocolitis. *Gut* (1996) 39(4):605–8. doi:10.1136/gut.39.4.605
  62. Hajishengallis G, Moutsopoulos NM. Role of bacteria in leukocyte adhesion deficiency-associated periodontitis. *Microb Pathog* (2016) 94:21–6. doi:10.1016/j.micpath.2015.09.003
  63. Savinko TS, Morrison VL, Uotila LM, Wolff CHJ, Alenius HT, Fagerholm SC. Functional beta2-integrins restrict skin inflammation in vivo. *J Invest Dermatol* (2015) 135(9):2249–57. doi:10.1038/jid.2015.164
  64. Lämmermann T, Bader BL, Monkley SJ, Words T, Wedlich-Söldner R, Hirsch K, et al. Rapid leukocyte migration by integrin-independent flowing and squeezing. *Nature* (2008) 453(7191):51–5. doi:10.1038/nature06887
  65. Chuluyan HE, Issekutz AC. VLA-4 integrin can mediate CD11/CD18-independent transendothelial migration of human monocytes. *J Clin Invest* (1993) 92(6):2768. doi:10.1172/JCI116895
  66. Henderson RB, Hobbs JA, Mathies M, Hogg N. Rapid recruitment of inflammatory monocytes is independent of neutrophil migration. *Blood* (2003) 102(1):328–35. doi:10.1182/blood-2002-10-3228
  67. Morrison VL, James MJ, Grzes K, Cook P, Glass DG, Savinko T, et al. Loss of beta2-integrin-mediated cytoskeletal linkage reprograms dendritic cells to a mature migratory phenotype. *Nat Commun* (2014) 5:5359. doi:10.1038/ncomms6359
  68. Benvenuti F. The dendritic cell synapse: a life dedicated to T cell activation. *Front Immunol* (2016) 7:70. doi:10.3389/fimmu.2016.00070
  69. Grakoui A, Bromley SK, Sumen C, Davis MM, Shaw AS, Allen PM, et al. The immunological synapse: a molecular machine controlling T cell activation. *Science* (1999) 285(5425):221–7. doi:10.1126/science.285.5425.221
  70. Van Severen GA, Shimizu Y, Horgan KJ, Shaw S. The LFA-1 ligand ICAM-1 provides an important costimulatory signal for T cell receptor-mediated activation of resting T cells. *J Immunol* (1990) 144(12):4579–86.
  71. Varga G, Balkow S, Wild MK, Stadtbauer A, Krummen M, Rothoef T, et al. Active MAC-1 (CD11b/CD18) on DCs inhibits full T-cell activation. *Blood* (2007) 109(2):661–9. doi:10.1182/blood-2005-12-023044
  72. Balkow S, Heinz S, Schmidbauer P, Kolanus W, Holzmann B, Grabbe S, et al. LFA-1 activity state on dendritic cells regulates contact duration with T cells and promotes T-cell priming. *Blood* (2010) 116(11):1885–94. doi:10.1182/blood-2009-05-224428
  73. Podgrabska S, Kamalu O, Mayer L, Shimaoka M, Snoeck H, Randolph GJ, et al. Inflamed lymphatic endothelium suppresses dendritic cell maturation and function via Mac-1/ICAM-1-dependent mechanism. *J Immunol* (2009) 183(3):1767–79. doi:10.4049/jimmunol.0802167
  74. Behrens EM, Sriram U, Shivers DK, Gallucci M, Ma Z, Finkel TH, et al. Complement receptor 3 ligation of dendritic cells suppresses their stimulatory capacity. *J Immunol* (2007) 178(10):6268–79. doi:10.4049/jimmunol.178.10.6268
  75. Ehrichtiou D, Xiong Y, Xu G, Chen W, Shi Y, Zhang L. CD11b facilitates the development of peripheral tolerance by suppressing Th17 differentiation. *J Exp Med* (2007) 204(7):1519–24. doi:10.1084/jem.20062292
  76. Ingalls RR, Golenbock DT. CD11c/CD18, a transmembrane signaling receptor for lipopolysaccharide. *J Exp Med* (1995) 181(4):1473–9. doi:10.1084/jem.181.4.1473
  77. Rezzonico R, Imbert V, Chicheportiche R, Dayer JM. Ligation of CD11b and CD11c  $\beta_2$  integrins by antibodies or soluble CD23 induces macrophage inflammatory protein 1 $\alpha$  (MIP-1 $\alpha$ ) and MIP-1 $\beta$  production in primary human monocytes through a pathway dependent on nuclear factor- $\kappa$ B. *Blood* (2001) 97(10):2932–40. doi:10.1182/blood.V97.10.2932
  78. Ling GS, Bennett J, Woollard KJ, Szajna M, Fossati-Jimack L, Taylor PR, et al. Integrin CD11b positively regulates TLR4-induced signalling pathways in dendritic cells but not in macrophages. *Nat Commun* (2014) 5:3039. doi:10.1038/ncomms4039
  79. Fan ST, Edgington TS. Coupling of the adhesive receptor CD11b/CD18 to functional enhancement of effector macrophage tissue factor response. *J Clin Invest* (1991) 87(1):50. doi:10.1172/JCI115000
  80. Ingalls RR, Arnaout MA, Golenbock DT. Outside-in signaling by lipopolysaccharide through a tailless integrin. *J Immunol* (1997) 159(1):433–8.
  81. Perera PY, Mayadas TN, Takeuchi O, Akira S, Zaks-Zilberman M, Goyert SM, et al. CD11b/CD18 acts in concert with CD14 and toll-like receptor (TLR) 4 to elicit full lipopolysaccharide and taxol-inducible gene expression. *J Immunol* (2001) 166(1):574–81. doi:10.4049/jimmunol.166.1.574
  82. Yee NK, Hamerman JA.  $\beta_2$  integrins inhibit TLR responses by regulating NF- $\kappa$ B pathway and p38 MAPK activation. *Eur J Immunol* (2013) 43(3):779–92. doi:10.1002/eji.201242550
  83. Han C, Jin J, Xu S, Liu H, Li N, Cao X. Integrin CD11b negatively regulates TLR-triggered inflammatory responses by activating Syk and promoting degradation of MyD88 and TRIF via Cbl-b. *Nat Immunol* (2010) 11(8):734–42. doi:10.1038/ni.1908
  84. Wang L, Gordon RA, Huynh L, Su X, Park Min KH, Han J, et al. Indirect inhibition of toll-like receptor and type I interferon responses by ITAM-coupled receptors and integrins. *Immunity* (2010) 32(4):518–30. doi:10.1016/j.immuni.2010.03.014
  85. Bai Y, Qian C, Qian L, Ma F, Hou J, Chen Y, et al. Integrin CD11b negatively regulates TLR9-triggered dendritic cell cross-priming by upregulating microRNA-146a. *J Immunol* (2012) 188(11):5293–302. doi:10.4049/jimmunol.1102371
  86. Kushchayeva Y, Mishchuk D, Ugarova T. The role of beta 2 integrins in macrophage migration during resolution of inflammation. *Blood* (2009) 114:3600.
  87. Nath SK, Han S, Kim-Howard X, Kelly JA, Viswanathan P, Gilkeson GS, et al. A nonsynonymous functional variant in integrin- $\alpha$ M (encoded by ITGAM) is associated with systemic lupus erythematosus. *Nat Genet* (2008) 40(2):152–4. doi:10.1038/ng.71
  88. Remmers EF, Plenge RM, Lee AT, Graham RR, Hom G, Behrens TW, et al. STAT4 and the risk of rheumatoid arthritis and systemic lupus erythematosus. *N Engl J Med* (2007) 357(10):977–86. doi:10.1056/NEJMoa073003
  89. de Lange KM, Moutsianas L, Lee JC, Lamb CA, Luo Y, Kennedy NA, et al. Genome-wide association study implicates immune activation of multiple integrin genes in inflammatory bowel disease. *Nat Genet* (2017) 49(2):256–61. doi:10.1038/ng.3760
  90. Thomas AP, Dunn TN, Oort PJ, Grino M, Adams SH. Inflammatory phenotyping identifies CD11d as a gene markedly induced in white adipose tissue in obese rodents and women. *J Nutr* (2011) 141(6):1172–80. doi:10.3945/jn.110.127068



91. Dinarello CA. Blocking interleukin-1 $\beta$  in acute and chronic autoinflammatory diseases. *J Intern Med* (2011) 269(1):16–28. doi:10.1111/j.1365-2796.2010.02313.x
92. Cush JJ, Lipsky PE. Phenotypic analysis of synovial tissue and peripheral blood lymphocytes isolated from patients with rheumatoid arthritis. *Arthritis Rheum* (1988) 31(10):1230–8. doi:10.1002/art.1780311003
93. Takahashi H, Söderström K, Nilsson E, Kiessling R, Patarroyo M. Integrins and other adhesion molecules on lymphocytes from synovial fluid and peripheral blood of rheumatoid arthritis patients. *Eur J Immunol* (1992) 22(11):2879–85. doi:10.1002/eji.1830221119
94. Jasin HE, Lightfoot E, Davis LS, Rothlein R, Faanes RB, Lipsky PE. Amelioration of antigen-induced arthritis in rabbits treated with monoclonal antibodies to leukocyte adhesion molecules. *Arthritis Rheum* (1992) 35(5):541–9. doi:10.1002/art.1780350508
95. Watts GM, Beurskens FJ, Martin-Padura I, Ballantyne CM, Klickstein LB, Brenner MB, et al. Manifestations of inflammatory arthritis are critically dependent on LFA-1. *J Immunol* (2005) 174(6):3668–75. doi:10.4049/jimmunol.174.6.3668
96. Suchard SJ, Stetsko DK, Davis PM, Skala S, Potin D, Launay M, et al. An LFA-1 ( $\alpha$ L $\beta$ 2) small-molecule antagonist reduces inflammation and joint destruction in murine models of arthritis. *J Immunol* (2010) 184(7):3917–26. doi:10.4049/jimmunol.0901095
97. Kakimoto K, Nakamura T, Ishii K, Takashi T, Iigou H, Yagita H, et al. The effect of anti-adhesion molecule antibody on the development of collagen-induced arthritis. *Cell Immunol* (1992) 142(2):326–37. doi:10.1016/0008-8749(92)90294-Y
98. Bullard DC, Hurley LA, Lorenzo I, Sly LM, Beaudet AL, Staite ND. Reduced susceptibility to collagen-induced arthritis in mice deficient in intercellular adhesion molecule-1. *J Immunol* (1996) 157(7):3153–8.
99. Stevanin M, Busso N, Chobaz V, Pigni M, Ghassem-Zadeh S, Zhang L, et al. CD11b regulates the Treg/Th17 balance in murine arthritis via IL-6. *Eur J Immunol* (2017) 47(4):637–45. doi:10.1002/eji.201646565
100. Taylor PC, Chu CQ, Plater-Zyberk C, Maini RN. Transfer of type II collagen-induced arthritis from DBA/1 to severe combined immunodeficiency mice can be prevented by blockade of Mac-1. *Immunology* (1996) 88(2):315–21. doi:10.1111/j.1365-2567.1996.tb00021.x
101. Leonardi CL, Papp KA, Gordon KB, Menter A, Feldman SR, Caro I, et al. Extended efalizumab therapy improves chronic plaque psoriasis: results from a randomized phase III trial. *J Am Acad Dermatol* (2005) 52(3):425–33. doi:10.1016/j.jaad.2004.09.029
102. Major EO. Progressive multifocal leukoencephalopathy in patients on immunomodulatory therapies. *Annu Rev Med* (2010) 61:35–47. doi:10.1146/annurev.med.080708.082655
103. Schwab N, Ulzheimer JC, Fox RJ, Schneider-Hohendorf T, Kieseier BC, Monoranu CM, et al. Fatal PML associated with efalizumab therapy insights into integrin  $\alpha$ L $\beta$ 2 in JC virus control. *Neurology* (2012) 78(7):458–67. doi:10.1212/WNL.0b013e3182478d4b
104. Pavlovic D, Patera AC, Nyberg F, Gerber M, Liu M; Progressive Multifocal Leukoencephalopathy Consortium. Progressive multifocal leukoencephalopathy: current treatment options and future perspectives. *Ther Adv Neurol Disord* (2015) 8(6):255–73. doi:10.1177/1756285615602832
105. Polman CH, O'Connor PW, Havrdova E, Hutchinson M, Kappos L, Miller DH, et al. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med* (2006) 354(9):899–910. doi:10.1056/NEJMoa044397
106. Ghosh S, Goldin E, Gordon FH, Malchow HA, Rask-Madsen J, Rutgeerts P, et al. Natalizumab for active Crohn's disease. *N Engl J Med* (2003) 348(1):24–32. doi:10.1056/NEJMoa020732
107. Berger JR, Fox RJ. Reassessing the risk of natalizumab-associated PML. *J Neurovirol* (2016) 22(4):533–5. doi:10.1007/s13365-016-0427-6
108. Kavanaugh AF, Davis LS, Jain RI, Nichols LA, Norris SH, Lipsky PE. A phase I/II open label study of the safety and efficacy of an anti-ICAM-1 (intercellular adhesion molecule-1; CD54) monoclonal antibody in early rheumatoid arthritis. *J Rheumatol* (1996) 23(8):1338–44.
109. Vuorte J, Lindsberg PJ, Kaste M, Meri S, Jansson SE, Rothlein R, et al. Anti-ICAM-1 monoclonal antibody R6.5 (enlimomab) promotes activation of neutrophils in whole blood. *J Immunol* (1999) 162(4):2353–7.
110. Grönholm M, Jahan F, Bryushkova EA, Madhavan S, Agliarolo F, Soto Hinojosa L, et al. LFA-1 integrin antibodies inhibit leukocyte  $\alpha$ 4 $\beta$ 1-mediated adhesion by intracellular signaling. *Blood* (2016) 128(9):1270–81. doi:10.1182/blood-2016-03-705160
111. Shimaoka M, Shifman JM, Jing H, Takagi J, Mayo SL, Springer TA. Computational design of an integrin I domain stabilized in the open high affinity conformation. *Nat Struct Mol Biol* (2000) 7(8):674–8. doi:10.1038/77978
112. Roberts AL, Fűrnröhr BG, Vyse TJ, Rhodes B. The complement receptor 3 (CD11b/CD18) agonist leukadherin-1 suppresses human innate inflammatory signalling. *Clin Exp Immunol* (2016) 185(3):361–71. doi:10.1111/cei.12803
113. Potin D, Launay M, Monatik F, Malabre P, Fabreguettes M, Fouquet A, et al. Discovery and development of 5-[(5S,9R)-9-(4-cyanophenyl)-3-(3,5-dichlorophenyl)-1-methyl-2, 4-dioxo-1, 3, 7-triazaspiro [4.4] non-7-yl-methyl]-3-thiophenecarboxylic acid (BMS-587101) a small molecule antagonist of leukocyte function associated antigen-1. *J Med Chem* (2006) 49(24):6946–9. doi:10.1021/jm0610806

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The handling Editor declared a shared affiliation, though no other collaboration, with the authors LS and CH.

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